

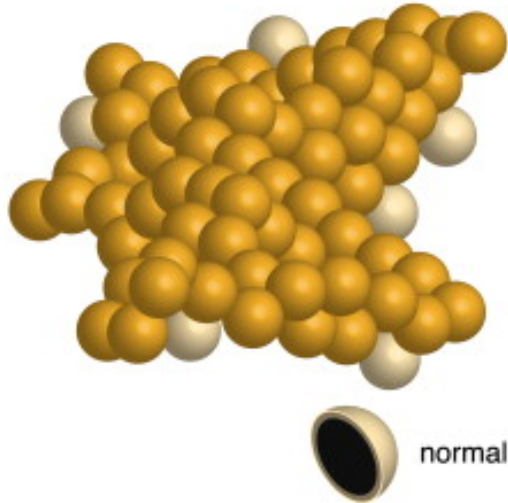


Assessing tumor heterogeneity and tracking clonal evolution using whole genome or exome sequencing

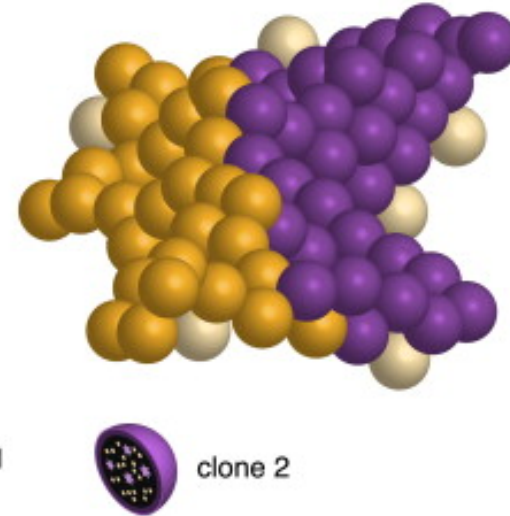
Chris Miller, PhD

Tumors are heterogeneous

A Schematic depiction of a mono-clonal tumor



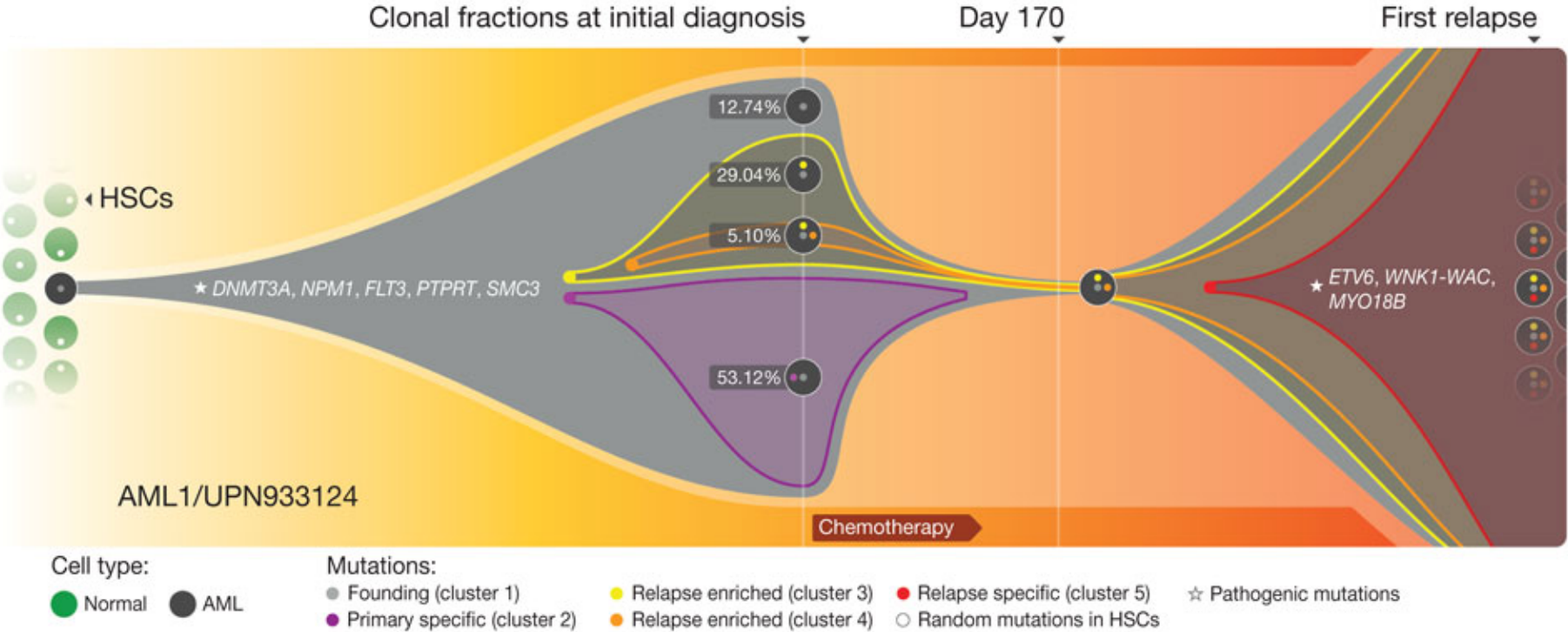
B Schematic depiction of a bi-clonal tumor



genetically diverse populations of cells
evolution occurs at the cellular level



Clonal evolution in relapsed AML

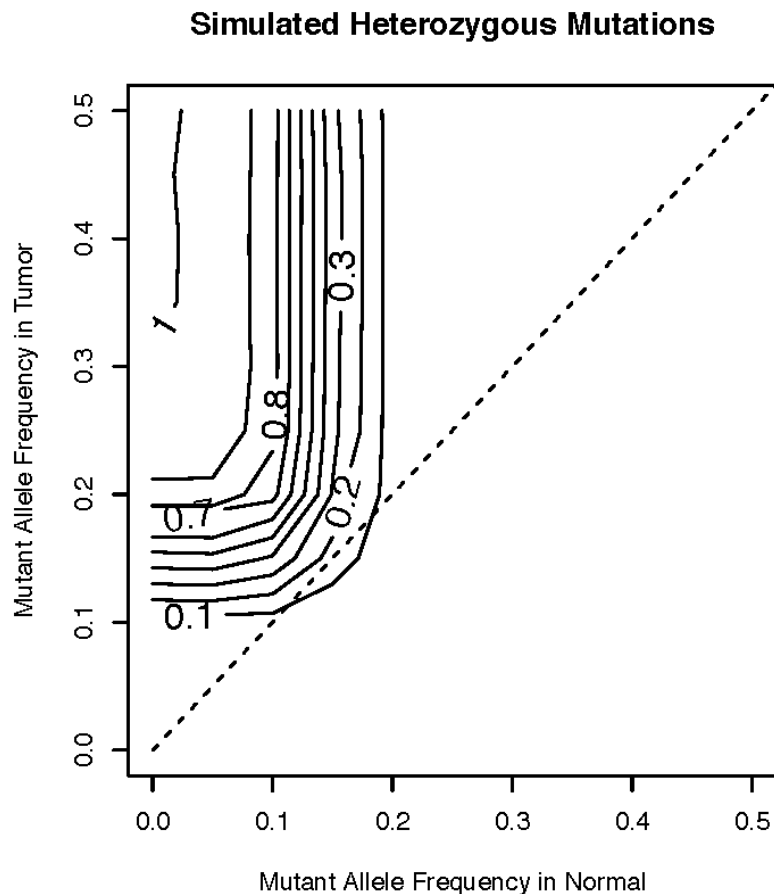


Challenges for detecting minor subclones

- Genomes are sequenced with low coverage
 - 30x not enough
- Algorithms aren't designed to detect low-frequency events



Somatic Sniper power simulations



- 90x coverage
- Power to detect event at 20% VAF: 85%
- Power to detect event at 10% VAF: 10%



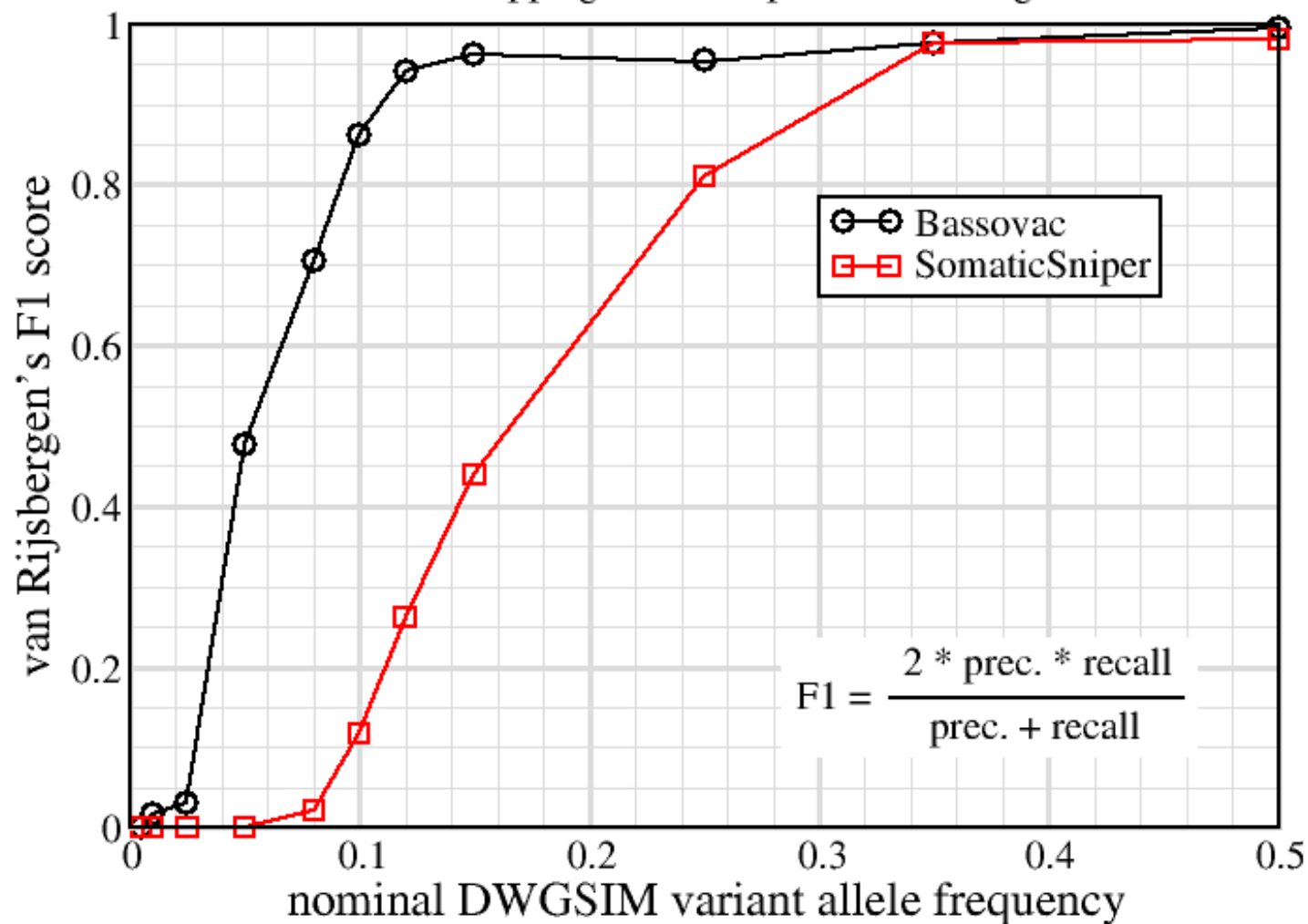
BASSOVAC

- **BA**yesian **S**coring of **S**omatic **V**ariant read **C**ounts
- Incorporates purity, ploidy, base quality, allele frequency, and overall mutation rate.
- Bayesian framework for inversion to obtain, probabilities of heterozygous and homozygous somatic events, given the data



Simulation

90X tumor 30X normal, Q20 simulation data, BMR 1e-6
with mapping and false-positive filtering



Real-world testing

- Primary breast tumor
- Matched normal
- 3 different metastases:
 - Spinal
 - Liver
 - Adrenal

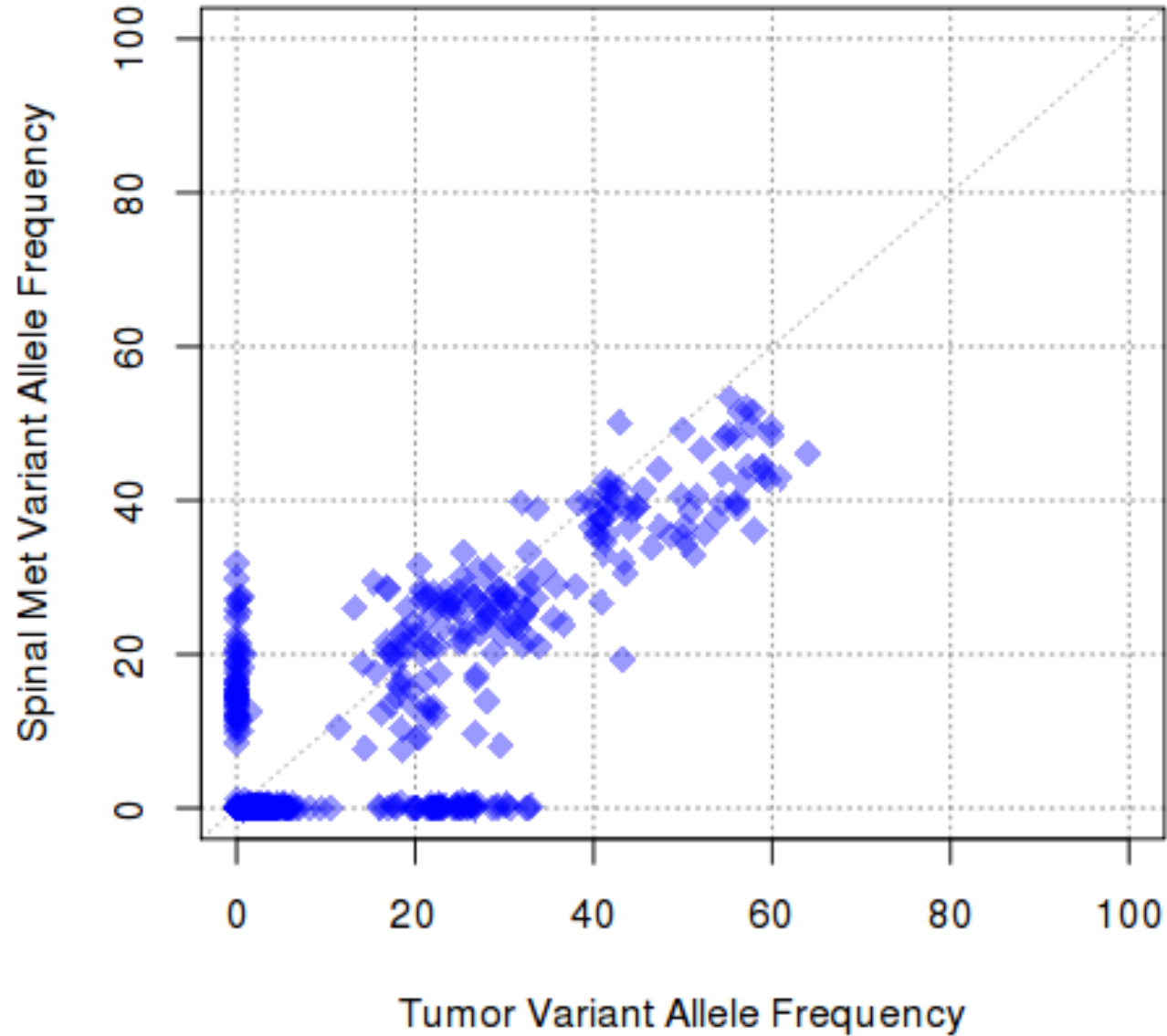


Real-world testing

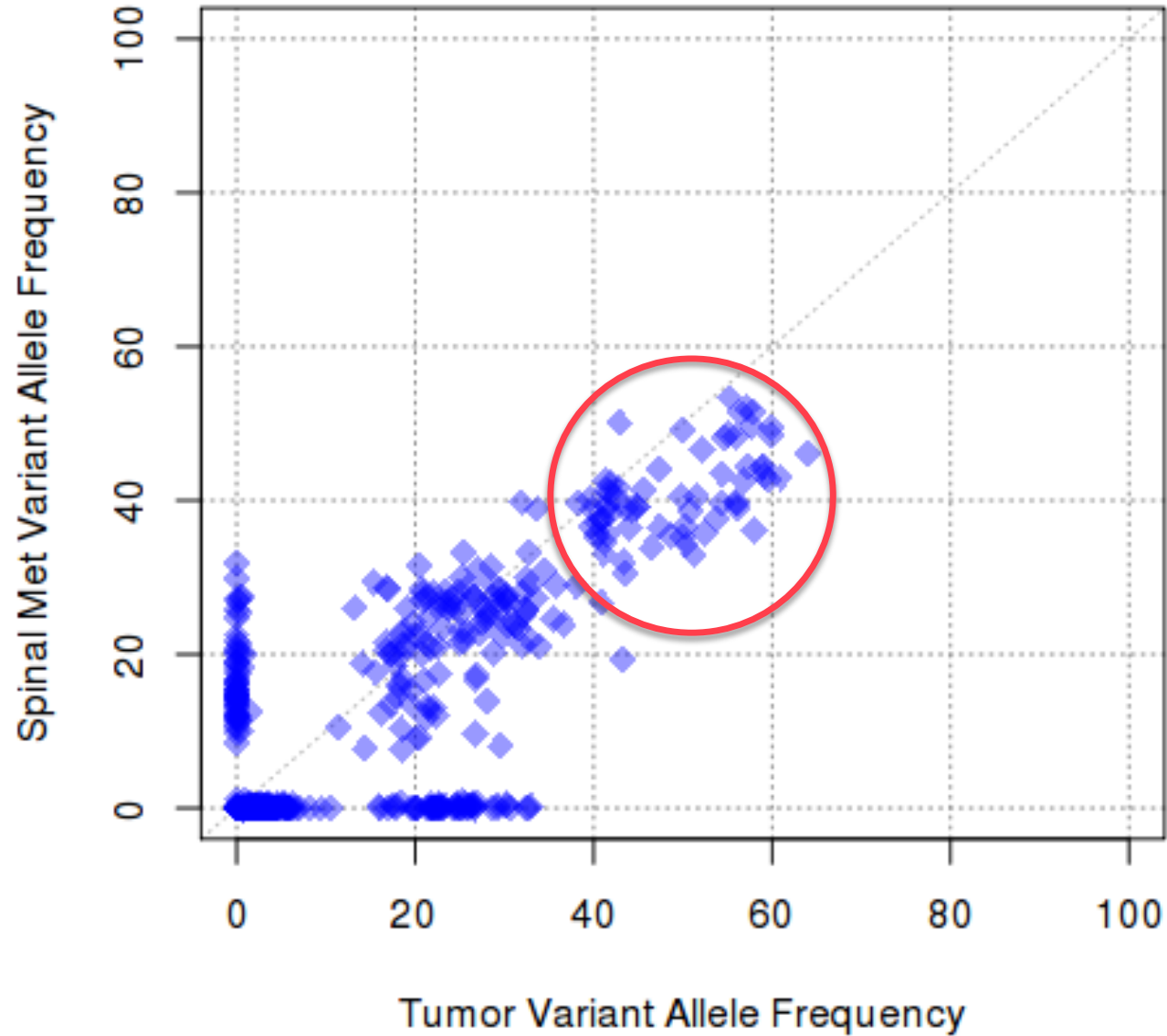
- All whole-genome sequenced to 30x
- Mutation calls made with Somatic Sniper and VarScan
- Capture validation performed for all variants
- Deep readcounts obtained from validation sequencing for all variants in all samples



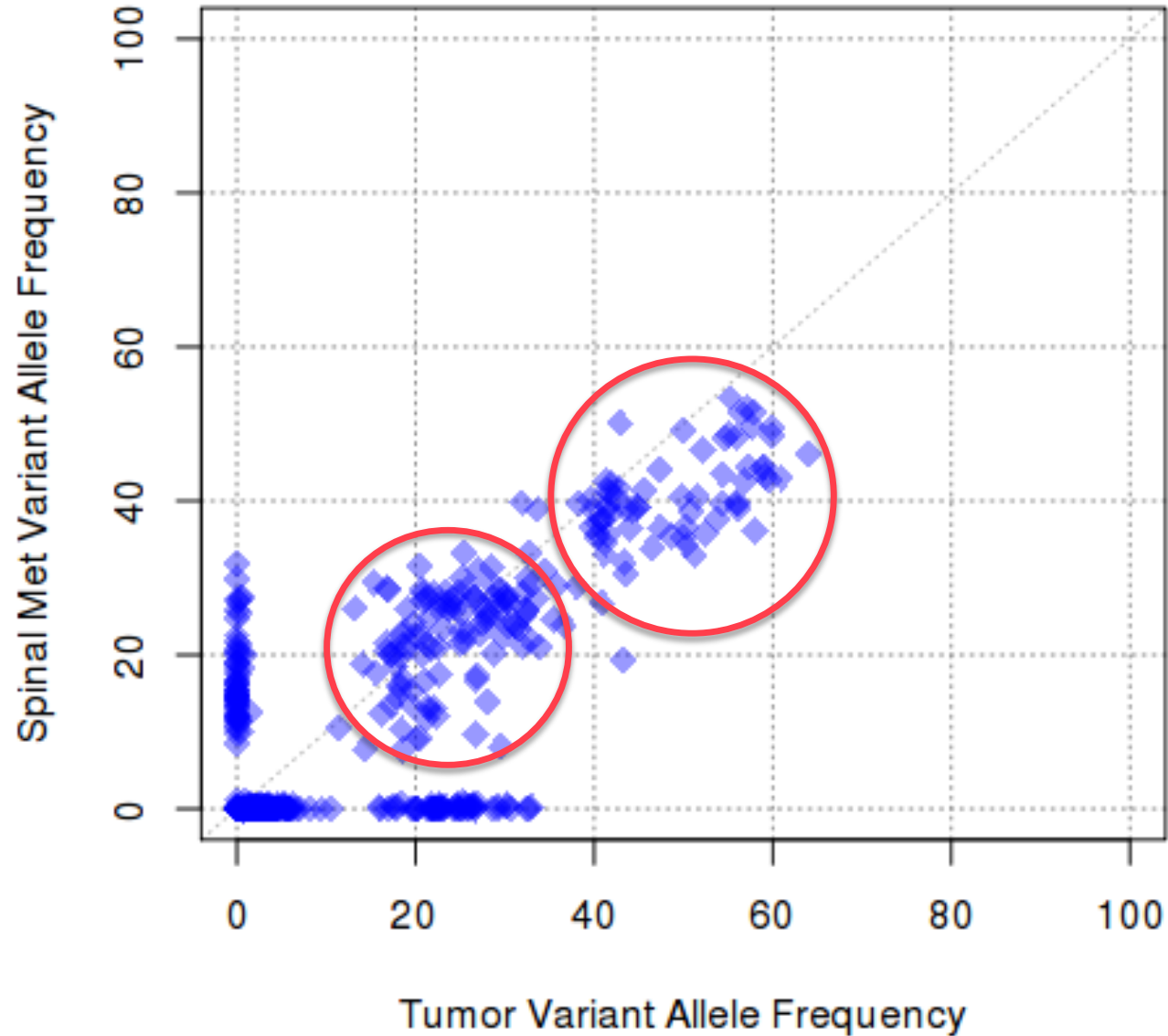
BRCA Tumor vs Spinal Metastasis



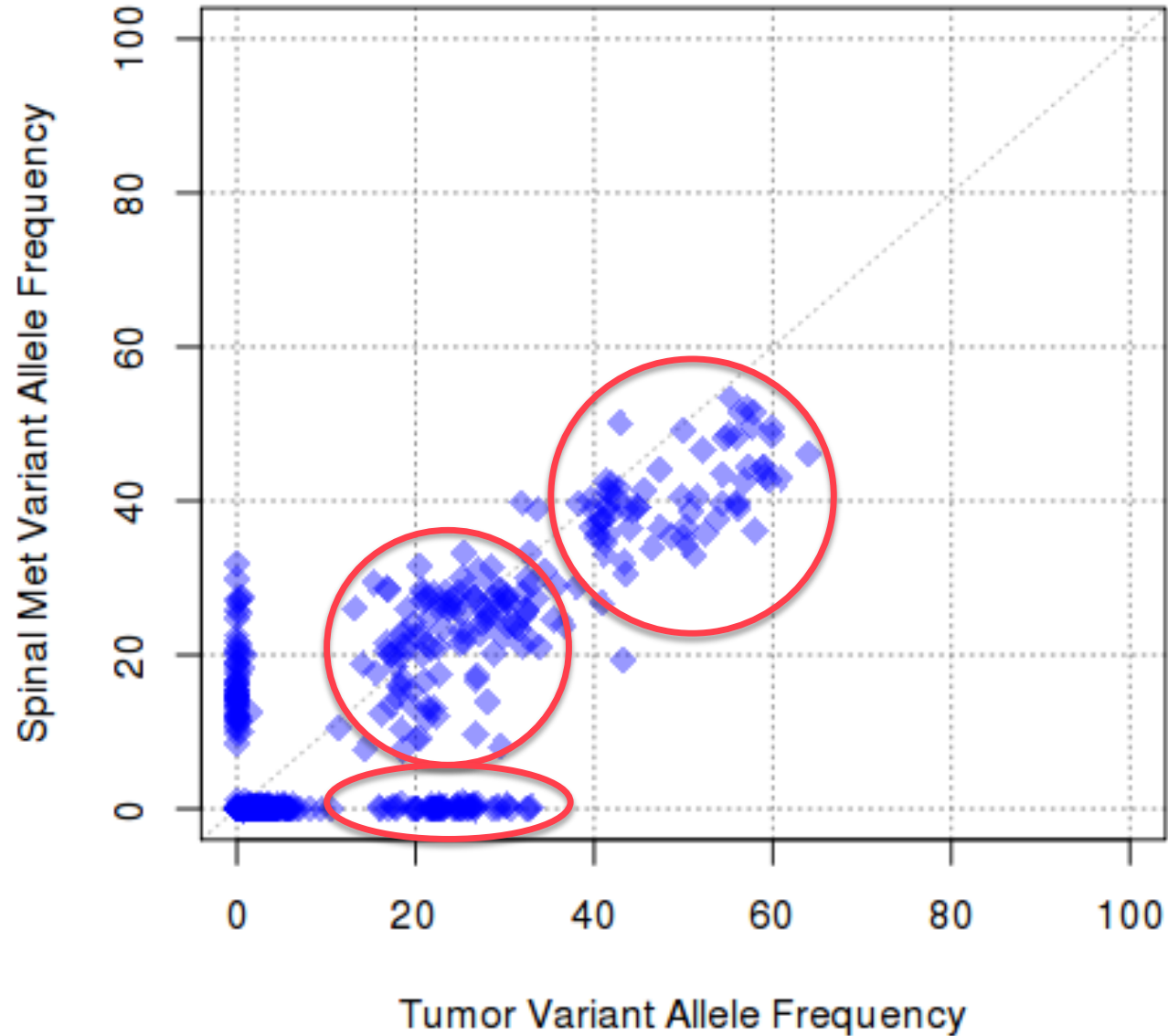
BRCA Tumor vs Spinal Metastasis



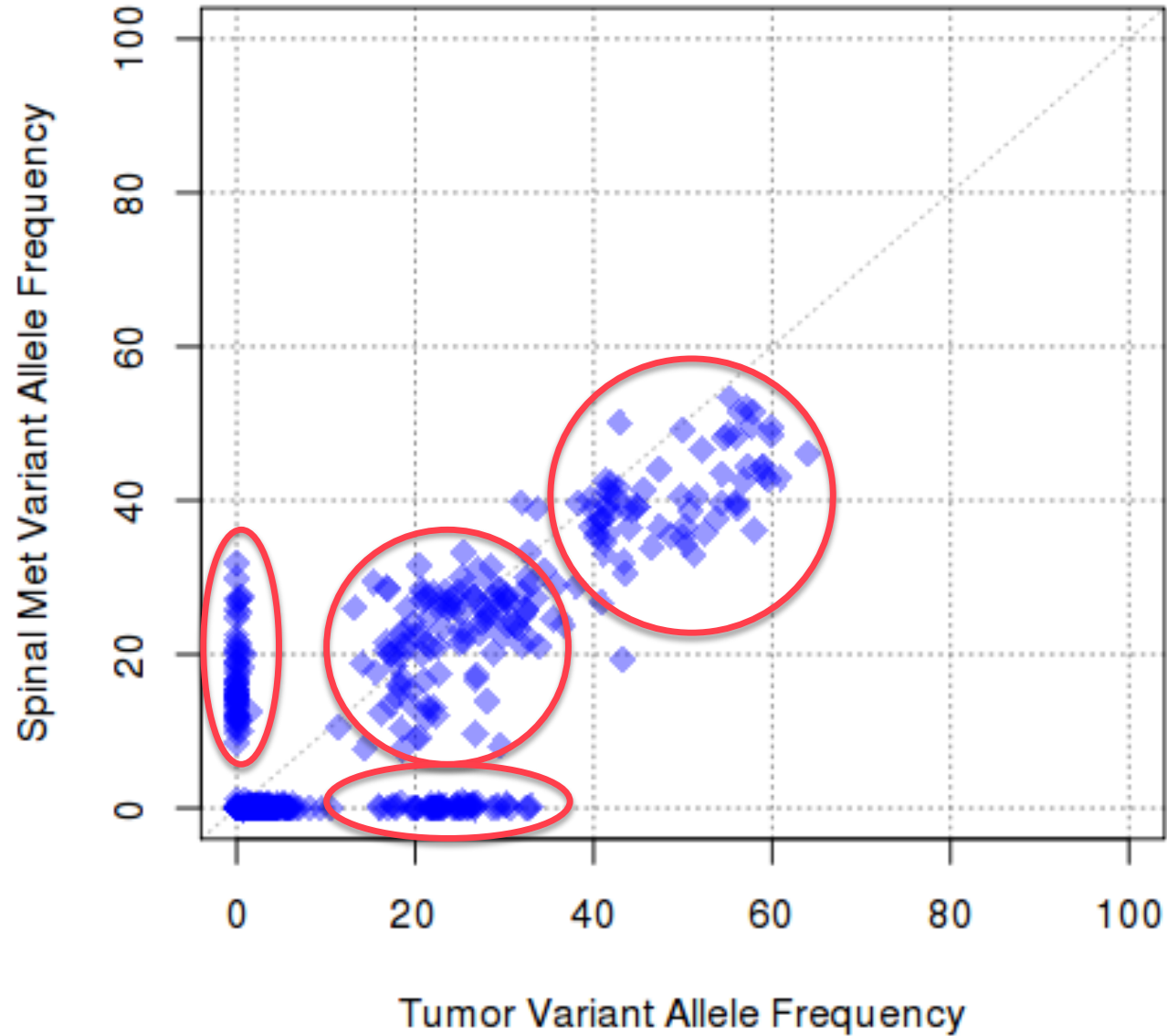
BRCA Tumor vs Spinal Metastasis



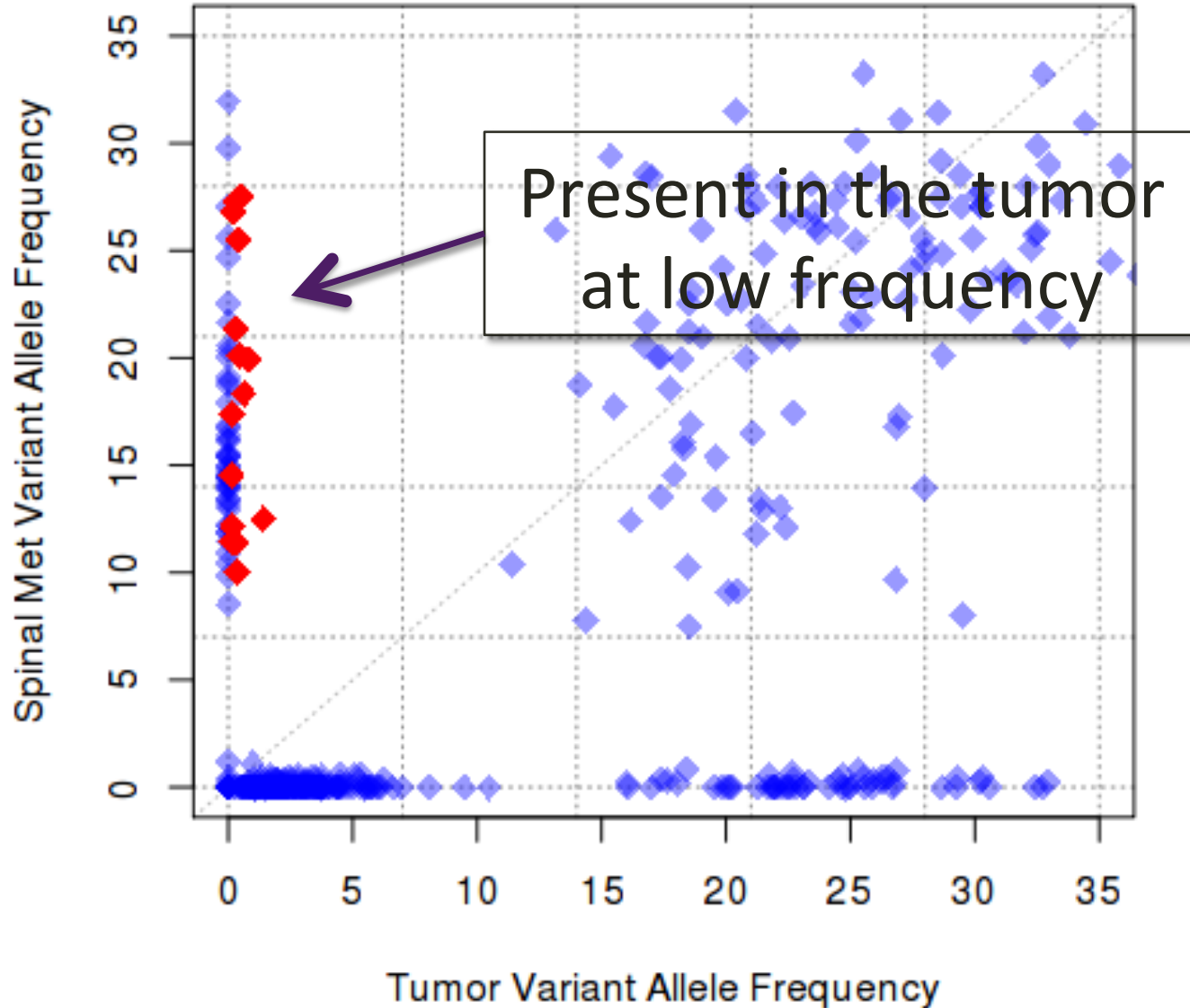
BRCA Tumor vs Spinal Metastasis



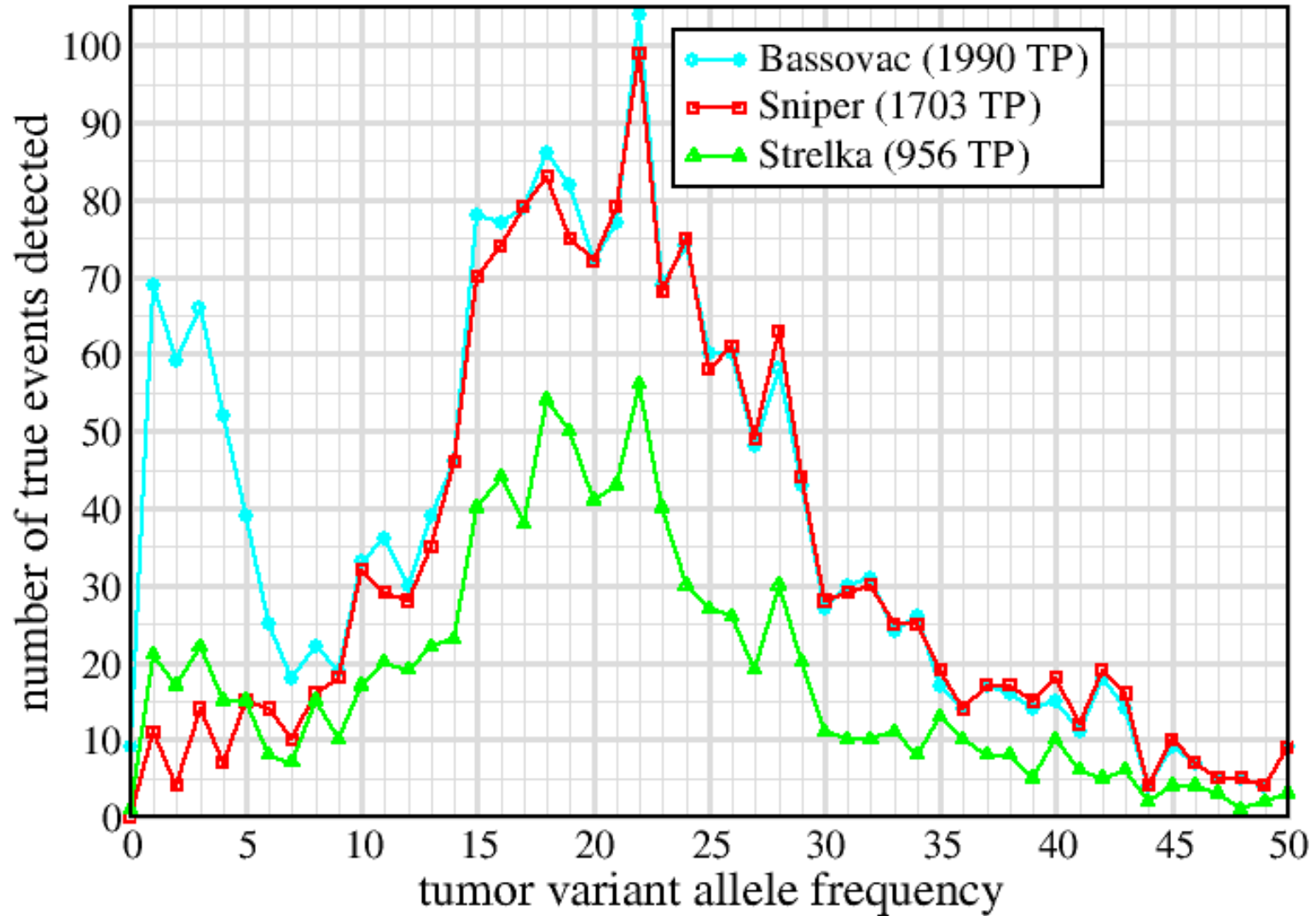
BRCA Tumor vs Spinal Metastasis



BRCA Tumor vs Spinal Metastasis



BASSOVAC sensitivity - BRCA met SNVs in tumor



BASSOVAC

- Over 50% of the variants present in the metastases are present at a detectable level in the tumor
- We can use BASSOVAC to detect true variants at very low frequencies ($< 2\%$)

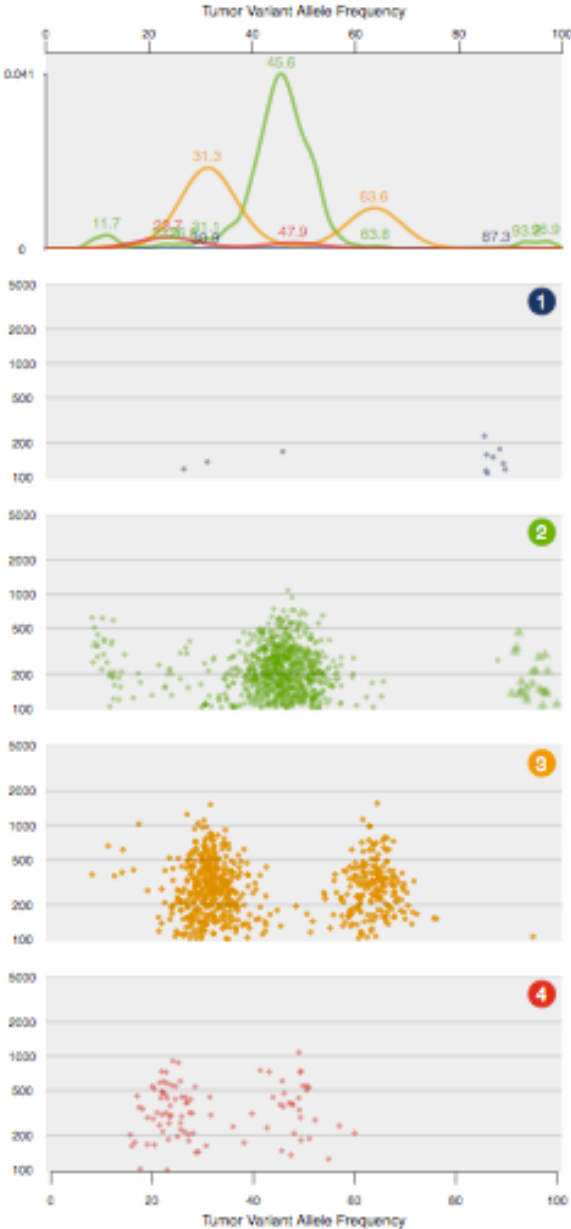


Clonal inference

- Given information about a tumor, how many clones are present?
 - Which variants are present in different subclones?
- Requires integrative approach
 - Variant allele frequencies
 - Copy number calls
 - Purity and Ploidy information

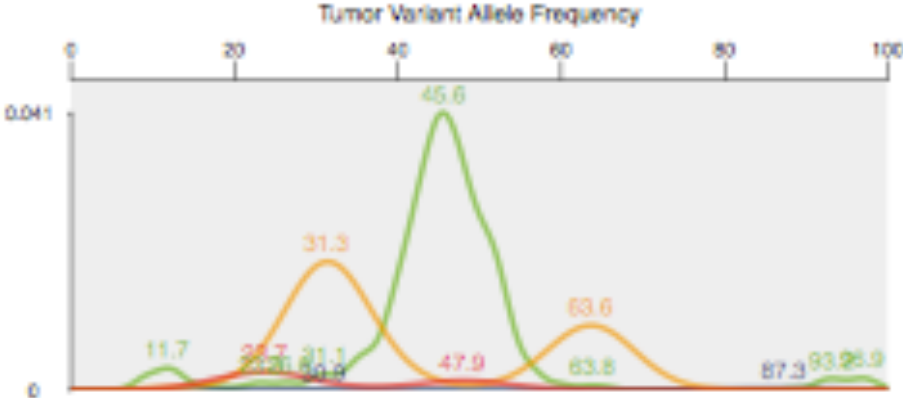


Clonality Plot

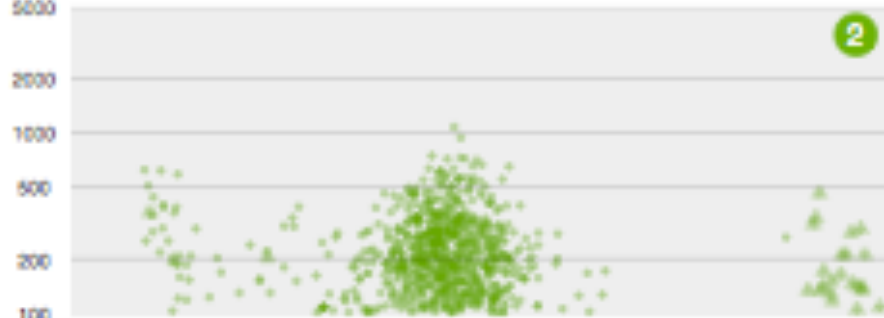


Clonality Plot

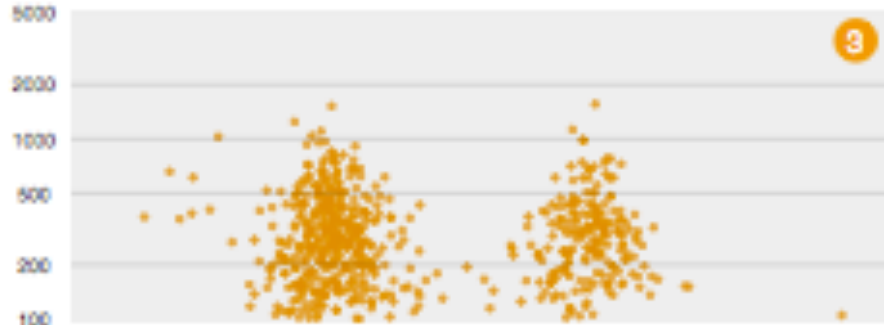
Kernel Density



SNVs in CN2 regions

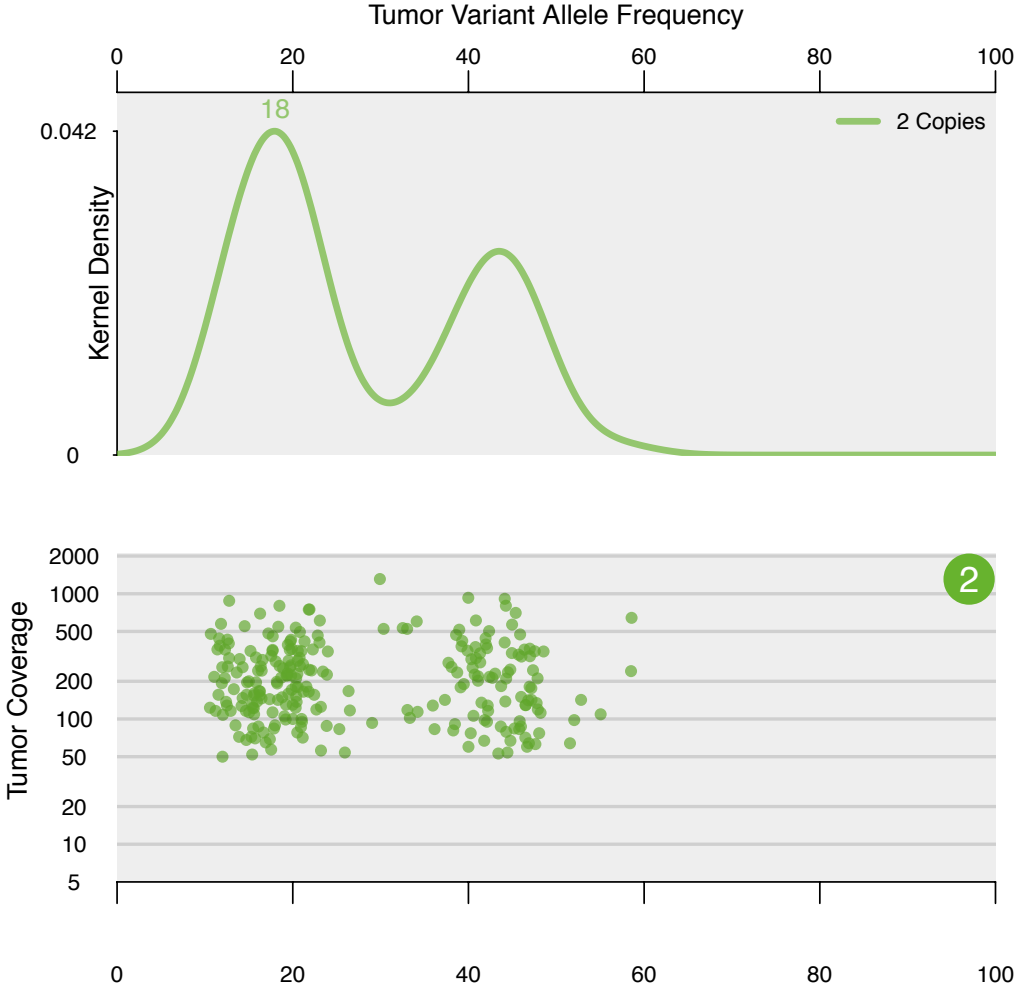


SNVs in CN3 regions



Clonality Plot

TCGA-B5-A0JV-01A-11D-A10B-09 Clonality Plot

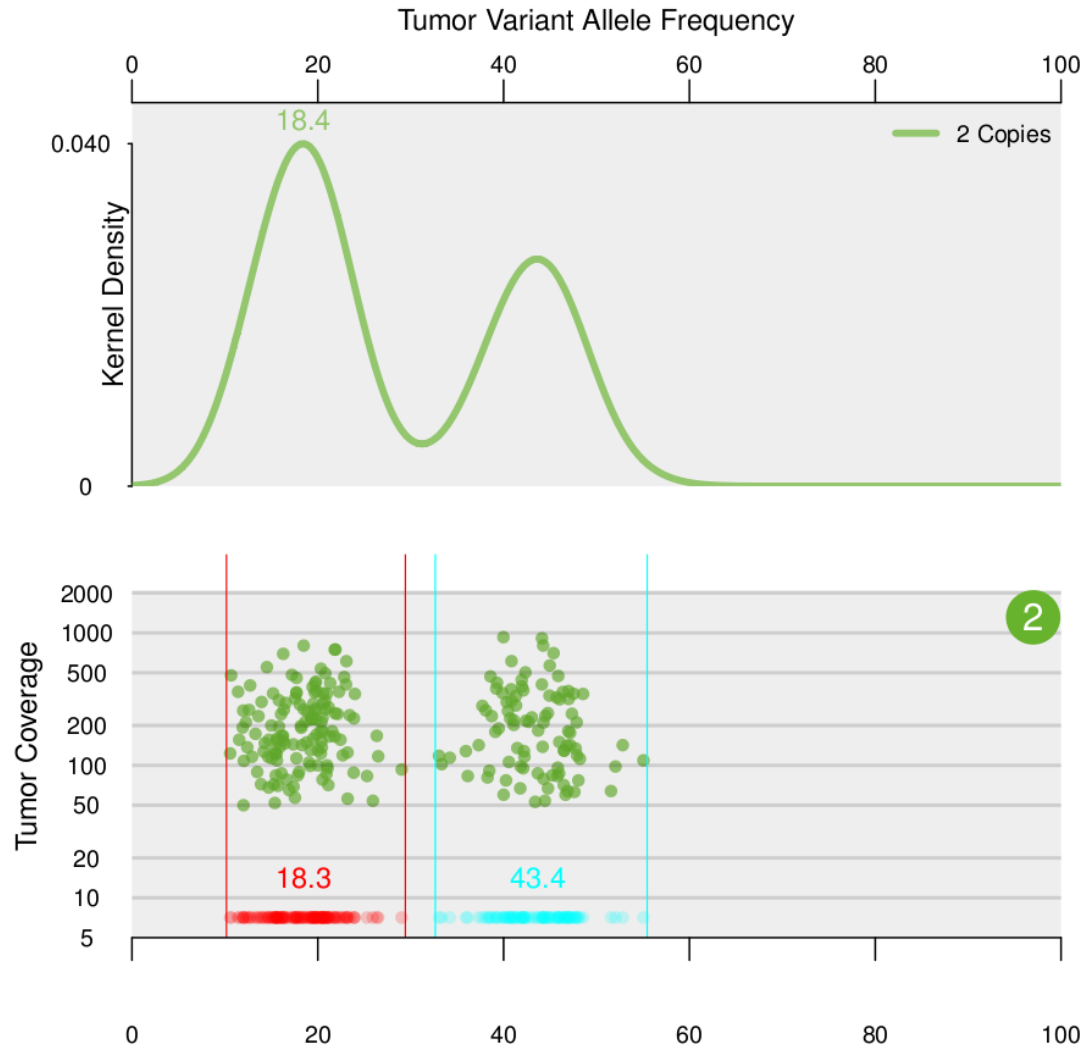


Infer clones in an automated, unbiased manner



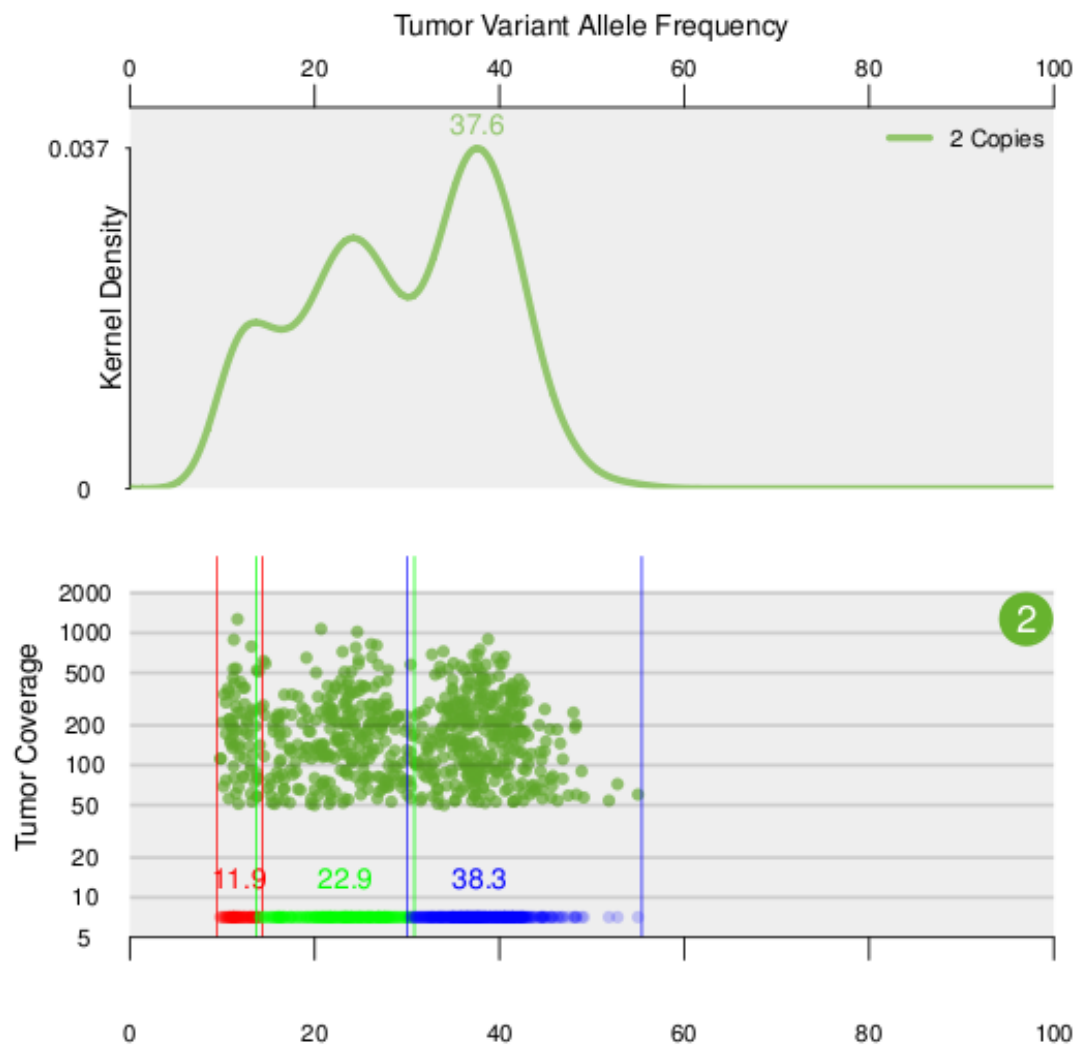
Biclonal sample

TCGA-B5-A0JV-01A-11D-A10B-09



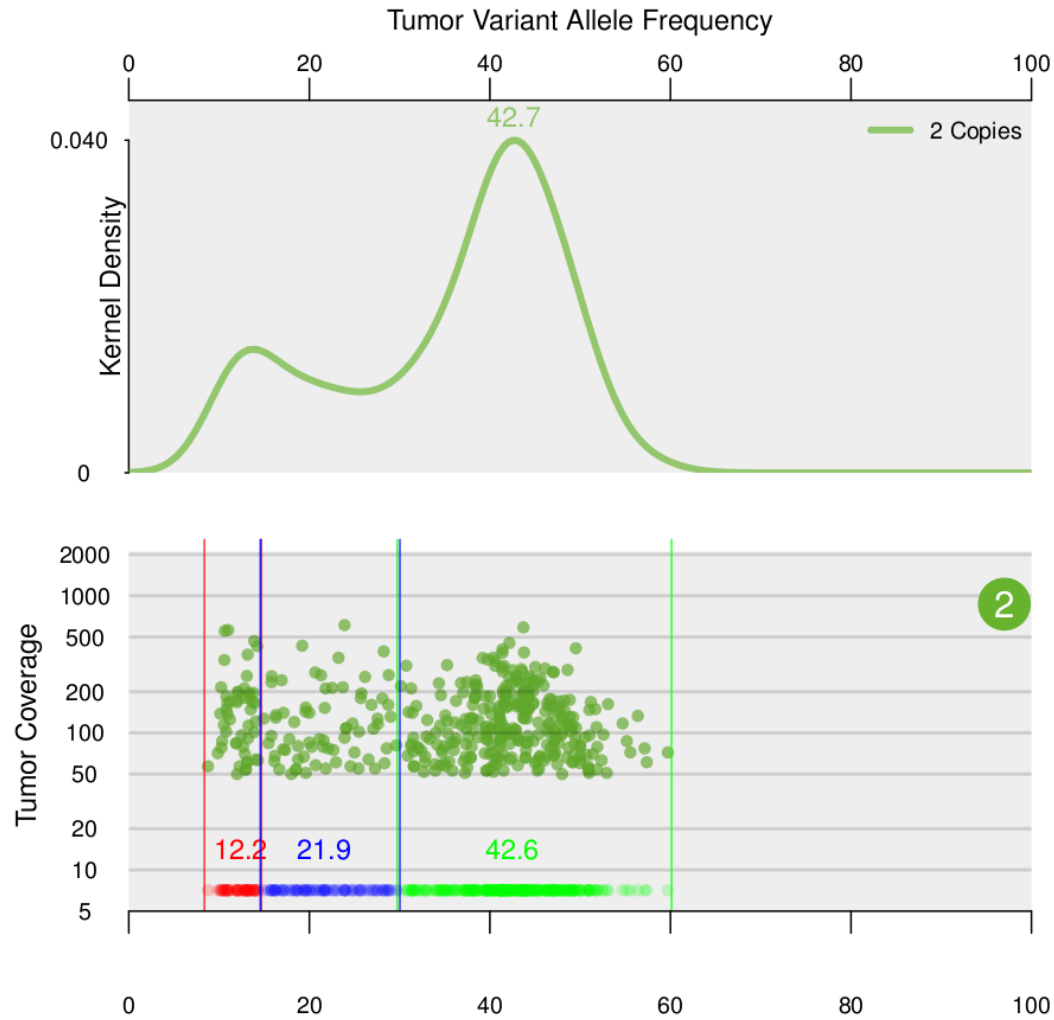
Triclonal sample

TCGA-AX-A063-01A-11W-A027-09



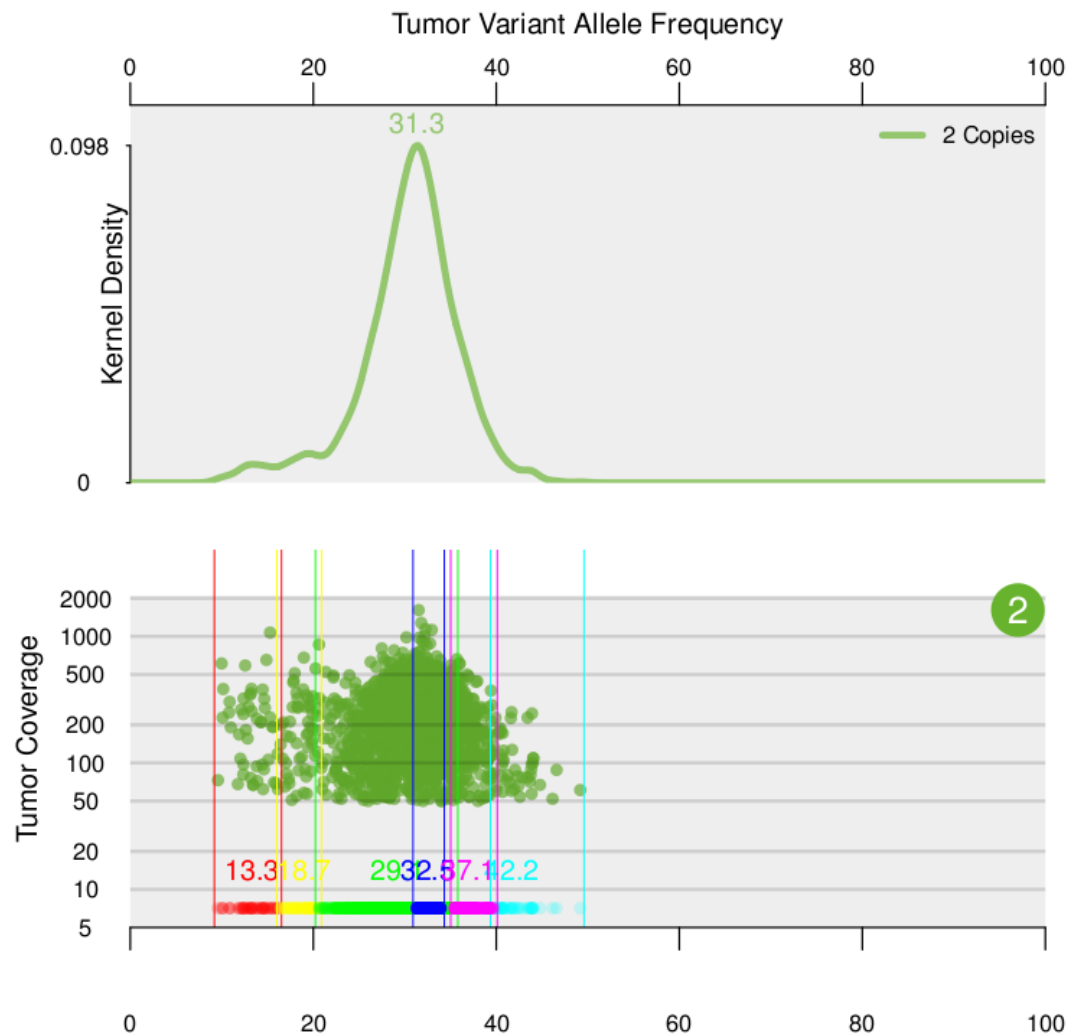
Non-intuitive sample

TCGA-BG-A0MQ-01A-11D-A10B-09



Multi-clonal Sample

TCGA-D1-A15X-01A-11D-A122-09



Clonal inference

- Most tumors have a founding clone and one or more subclones (LAML, BRCA, UCEC)
- Lower bound on number of clones



Conclusions

- We can detect somatic mutations at very low frequencies using BASSOVAC
- We have developed robust automatic methods for inferring details about the subclonal architecture of a tumor
- Goal: characterizing minor subclones at diagnosis, rather than discovering their presence at relapse



Acknowledgements

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