

# Technical and Logistical Approaches to Genomic Screening

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# Summary: Optimizing High Throughput Genomic Screening: Lessons Learned



Consistent DNA source and quality



Optimize and Simplify workflow with automation



Stable Process for Library Automation



Cost efficient



Continual upgrade process with testing, validation, implementation for both laboratory protocols and software upgrades



Metrics to produce a quality product and monitoring of production workflow



Semi-automated analysis pipeline



Clear communication of reporting practices

# General Requirements to Launch Clinical Test (LDT)

- Performed in CLIA certified laboratory, CAP accreditation
- Indication for testing, requisition, required elements
- Specimen requirements
- SOP, validated reagents, instruments, vendors
- Technical limitations determined and disclosed
- Assay interpretation
- Validation studies: Rationale used for validation
- Clinical reporting criteria
- Limitations of testing and reporting
- Post - launch evaluation

# Clinical Test Validation

## Analytical validation

- Accuracy
- Precision
- Sensitivity/Specificity
- Reproducibility
- Limit of Detection
- Choice of validation samples

## Clinical Validation

- Purpose of test
  - Newborn screening – actionable disorders of early childhood
  - Family planning – carrier screening
  - Wellness – hereditary cancer, CDC Tier 1, ACMG 59, PGX
- Evidence
- Actionability



# Stages of Validation of Clinical NGS Test

- Test Development and Optimization
  - Indication for testing, test scope, samples types accepted, TAT
  - Wet lab workflow, automation, QA metrics and performance
  - Establish analytical pipeline, versioning, additional modules for challenging regions
  - Variant confirmation approach
  - Validation strategy
- Test Validation
  - Measure test performance through analytical validation - sensitivity (recall, positive percent agreement), specificity (negative percent agreement), precision, reproducibility (comparison across instruments and operators), repeatability (technical replicates under same condition)
  - Stress test performance to determine limitations - difficult regions, mosaicism detection level
- Test Performance Monitoring / Quality Management
  - QA metrics established and tracked
    - Sample identity, contamination, GB>Q30, 20x%



# Sample Collection Considerations

- Participant convenience
- Kitting, detailed instructions for self-collection
- Bullet-proof labeling and downstream matching to contact information
- Cost
- Ability to automate DNA extraction
- Sample stability – during shipment and time to processing
- Failure rate of chosen method
- Potential ability to store DNA long-term (Biorepository)

# Sample Collection Options

## Saliva / Mouthwash/ Buccal

- 1 Read and sign the enclosed consent form.
- 2 Do NOT eat, drink, smoke or chew gum for 30 minutes before giving your saliva sample.
- 3 Do NOT remove the plastic film from the funnel lid that contains the clear liquid.

4



Spit into the open funnel until the amount of saliva (not bubbles) reaches the fill line. Most people take 2 to 5 minutes to fill the tube.

5



Close the lid tightly by pushing down hard on the funnel lid until you hear a loud click. The liquid will flow down into the tube.

6



Hold the tube upright. Unscrew the funnel lid from the tube and discard.

7



8




9



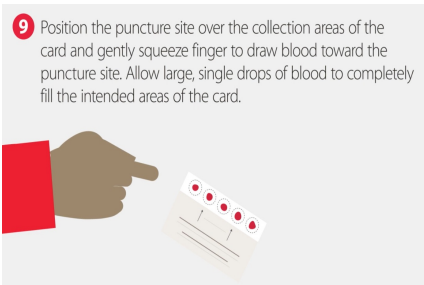
Source: www.cdc.gov/ncbddd

## Dried Blood Spot

- 6 Remove the lancet cap by twisting and pulling, then dispose. Lancet type may vary by test.

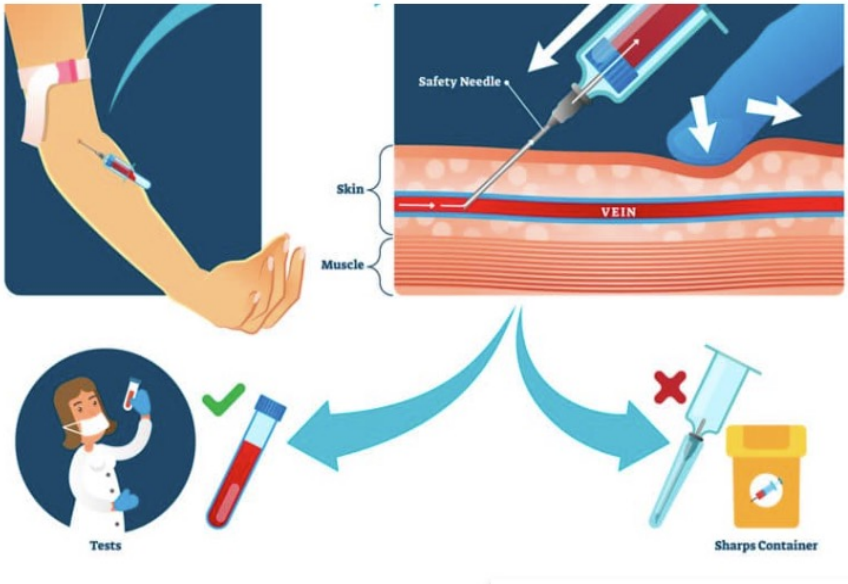


- 9 Position the puncture site over the collection areas of the card and gently squeeze finger to draw blood toward the puncture site. Allow large, single drops of blood to completely fill the intended areas of the card.



Source: Cardinal Health

## Whole Blood

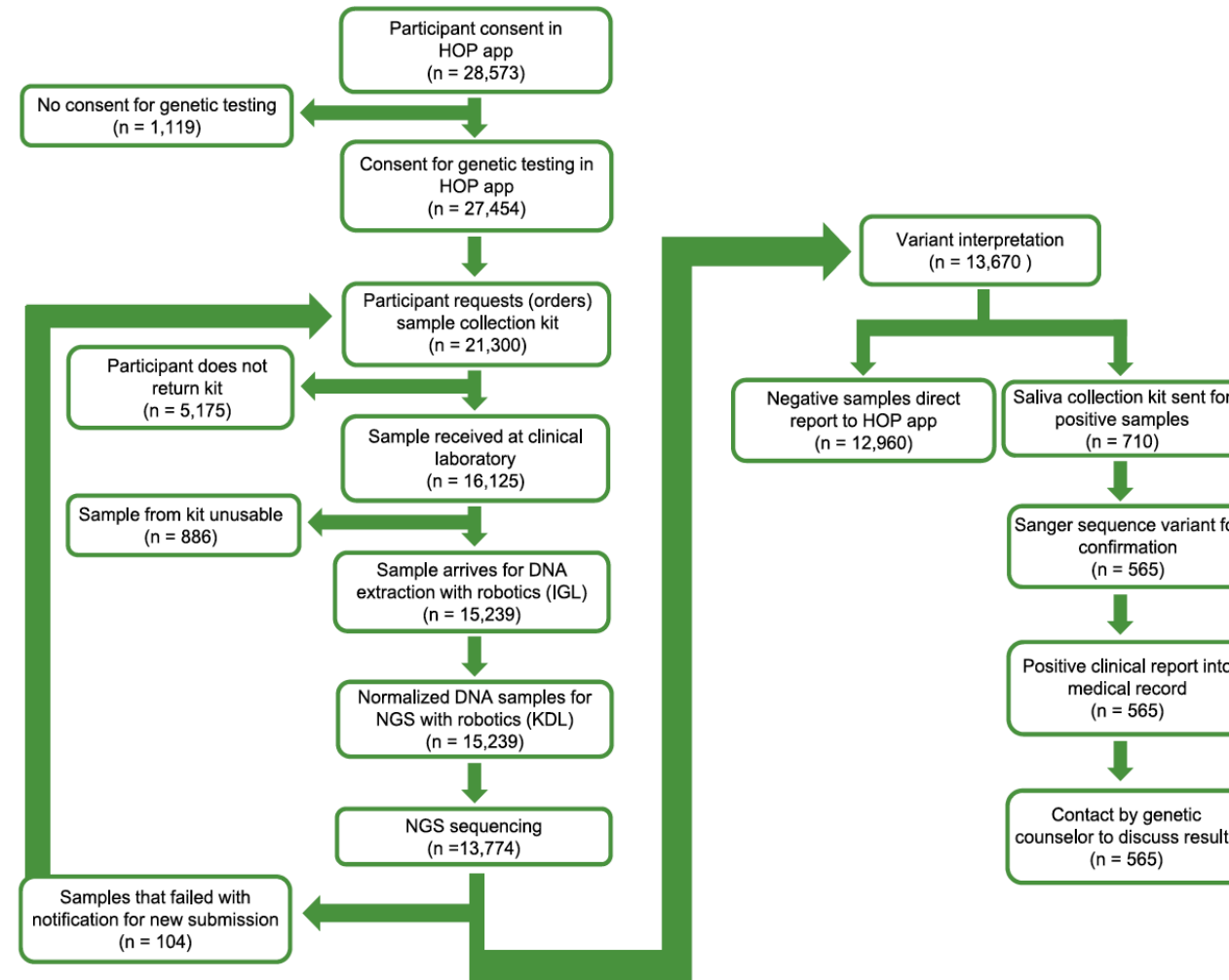


The diagram illustrates the process of drawing a whole blood sample. It shows a safety needle being inserted into a vein in the arm. Labels include 'Skin' and 'Muscle' for the layers of the arm, and 'VEIN' for the blood vessel. A 'Safety Needle' is shown with a red stopcock. Below the diagram, a nurse is shown holding a test tube, labeled 'Tests'. A green checkmark indicates a successful draw, while a red 'X' indicates an unsuccessful attempt. A 'Sharps Container' is shown for disposal.

Source: Arizona College of Nursing



# Sample Workflow



**Figure 1. Pipeline for the enrollment of HOP participants and sample workflow**

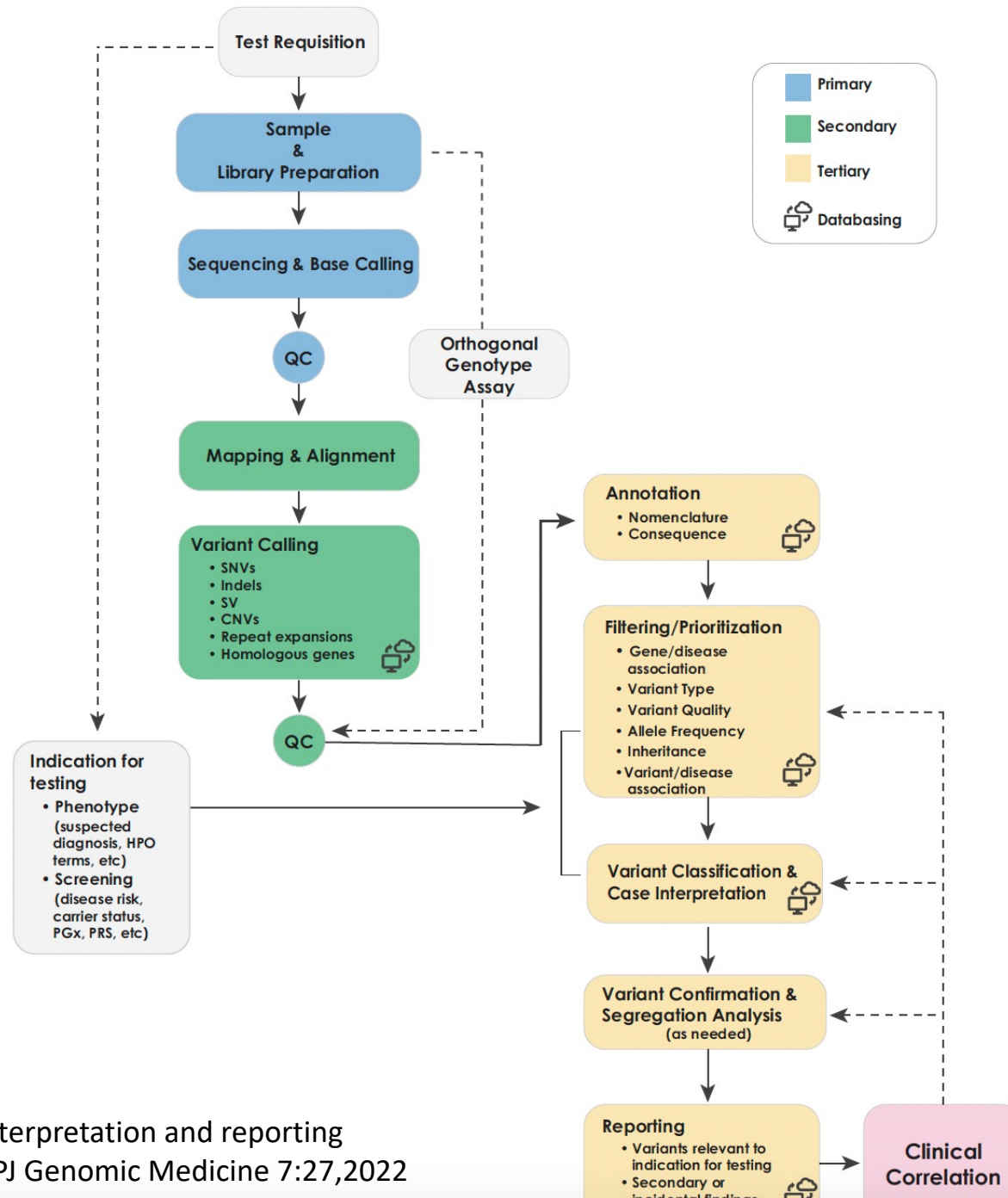
HOP participants sequenced as of April 15, 2022. Differences in numbers are reflective of the fluid pipeline and time it takes samples to process from consent in the HOP app through next-generation sequencing (NGS) and analysis. IGL, Integrated Genomics Laboratory, core laboratory; KDL, Knight Diagnostic Laboratories, clinical laboratory.



# Choice of Platform: Planning for High Throughput Testing

- Genotyping array
  - Example: UK Biobank Array - 800,000 markers
    - Exome, GWAS, PGX, HLA
    - Samples screened at Biobanks, genome centers, core labs
  - GDA array - All of US, ancestry, concordance
  - Arrays cost effective, high throughput, low failure rate, less flexibility after design
- Targeted NGS panel
  - Example: universal carrier screening panels, hereditary cancer panels
  - Less data produced, higher coverage, better CNV detection, less flexibility after design
- WES
  - Less data, less cost than WGS, ability to re-analyze, missing regions important for PGX, PRS
- WGS
  - Hypothesis-free, ability to re-analyze, highest cost and data
- Hybrid design - low pass WGS with 30-40X WES

# NGS Workflow



- Technical production
- Identification of DNA variants
- Annotation, prioritization, interpretation, reporting

# Analysis / Variant Classification

- Variant calling via bioinformatic pipeline
- Annotation, application of filters, database searches, prediction of functional effect, population frequency
  - clinical significance in ClinVar
  - classification in Human Gene Mutation Database (HGMD)
  - variant type, and population frequency
- Automated search against established database
- Manual curation of novel variants

# Quality Metrics

There are three main areas where QC can be applied to next-generation sequencing (NGS)

On the starting nucleic acid samples

After library preparation

- Insert size

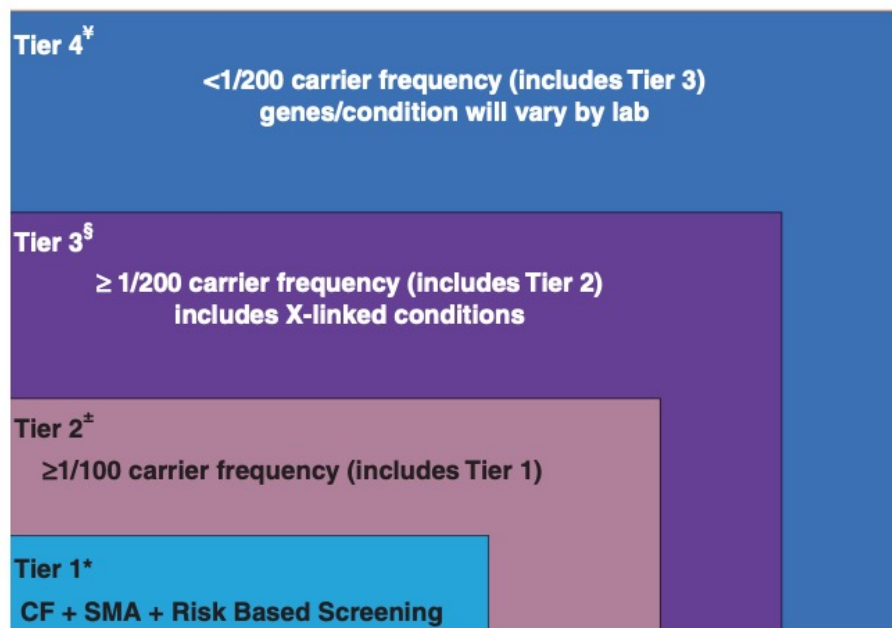
Post-sequencing (Pass/Fail)

- Sample identity and contamination
- Gb>Q30
- Mean coverage
- % Callability

Post Sequencing Monitoring

- Performance metrics
- Positive controls, periodic reference standard
- Periodic review of positivity rates (%positive calls by gene)

# Carrier Screening Panel Example



**Fig. 1 The Euler diagram shows an overlapping tiered approach to carrier screening.** \*Recommended by the American College of Medical Genetics and Genomics (ACMG)<sup>17,18</sup> and American College of Obstetricians and Gynecologists (ACOG).<sup>19</sup> <sup>±</sup>Recommended by ACOG.<sup>2</sup> <sup>§</sup>Supported by literature.<sup>49,50</sup> <sup>¶</sup>Offered by molecular testing laboratories; the list of genes/conditions may vary by the laboratory. *CF* cystic fibrosis, *SMA* spinal muscular atrophy.

## (Expanded Tier 4; 445 Gene) Carrier panel

- Exceeds current ACMG and ACOG recommendations
- Inherited X-linked carrier status identified for females
- 14 day TAT
- NGS panel with larger gene library than reported genes
- Detects SNV, indel, CNV, Path and LP variants reported
- Difficult to sequence genes (FMR1, FXN, CYP21A2) require separate assays and joining of data for reporting
- Orthogonal confirmations for specific genes

# WGS-Specific Validation and Reporting Considerations

- Determine which variant types will be reported
  - SNV, indel, CNV, SV, trinucleotide repeat, mitochondrial genome
  - Gather appropriate positive controls\* (common, rare)
  - Stress the system by selecting variants in challenging regions (high homology, pseudogenes)
  - Clearly define limits of detection
- Determine coverage level - depth of coverage, base quality, mapping quality
- Strategy for clinically important variants in difficult regions - additional bioinformatic modules, manual inspection of raw data, orthogonal methods, confirmation
- Tiered reporting
  - All of US - ancestry, hereditary disease risk, PGX
  - Subscription model

# Return of Results

- Critical to clearly communicate the result parameters to providers and participants
- Screening usually limited to Pathogenic or Likely Pathogenic
- Affected status (AD, AR, X-linked)
- Carrier status?
- Management guidance for positives
- Does negative result require counseling?
- Re Contact / Re Analysis / Subscription
  - Clear communication / expectation regarding “living report”

# Summary

- Overview of current state of clinical methods for population genomic screening
- Clear distinction between reporting of diagnostic vs screening genomic tests
- Laboratory quality measures are not distinct
- High throughput requirements can present unique challenges to the laboratory